Taxonomic Status of *Micrococcus agilis* Ali-Cohen 1889

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An amended description of *Micrococcus agilis* Ali-Cohen 1889 is proposed on the basis of a taxonomic study of five strains. Strain ATCC 966 (= CCM 2390 = NCTC 7509) is designated as the neotype strain of *M. agilis* Ali-Cohen 1889.

*Micrococcus agilis* Ali-Cohen 1889 is undoubtedly among the first described flagellated microccci. However, no type culture of *M. agilis* seems to exist in any strain collection. The strains labeled as *M. agilis* (ATCC 988 and 9814) maintained in the American Type Culture Collection were found to be nonmotile (13).

We received several strains of flagellated microccci that resemble *M. agilis* in their characteristics. The aim of the present paper is to give a revised description of *M. agilis* Ali-Cohen 1889 and to designate the neotype strain.

**MATERIALS AND METHODS**

**Organisms.** Five strains of aerobic, dark rose-red-pigmented microccci were investigated (Table 1). The strains were maintained on nutrient agar at 4°C. All morphological, nutritional, and biochemical studies were made on cultures grown at 25°C.

**Methods.** Morphological properties, Gram staining, starch hydrolysis, and nitrate reduction were determined according to methods described in the *Manual of Microbiological Methods* (4). Most of the cultural and biochemical methods were those described by Mortensen and Kocur (15). For glucose utilization, the medium recommended by the ICSB Subcommittee on Taxonomy of Staphylococci and Microccci was used (19). Gelatin hydrolysis was determined by the method of Clarke (3). Production of indole, hydrogen sulfide, acetoin, phenylalanine deaminase, and arginine dihydrolase, the methyl red test, and growth on Simmons’ citrate agar were studied by methods recommended by Ewing (6). Oxidase was detected by the method of Gaby and Hadley (7). Salt tolerance was observed on nutrient agar with 5, 7.5, 10, or 15% NaCl. Sensitivity to antibiotics was determined by the use of Oxoid multdisks.

The peptidoglycan type of the cell wall was determined by the method of Schleifer and Kandler (17).

**RESULTS AND DISCUSSION**

**Morphology.** All five strains examined were gram-positive cocci, 0.8 to 1.2 μm in diameter, arranged in pairs and in tetrads. Each cell was motile, with one to three flagella (Fig. 1). The only exception was strain CCM 2131, which was not flagellated.
gelatin hydrolysis, nitrate reduction, and production of acid from glucose. These discrepancies may be attributed to different techniques for the detection of the above test used by Ali-Cohen and us. In spite of these differences, we suggest that the five strains studied belong to *M. agilis*. An amended description of this species is given below.


(Figures in parenthesis are percentages of strains positive).

- Spheres 0.8 to 1.2 μm in diameter, occurring in pairs and tetrads. Motile by means of one or three flagella. Nonmotile strains may occur. Nonsporeforming. Gram positive.
- The cell wall peptidoglycan is of the L-Lys-L- Thr-L-Ala₄ type.
- Agar slant: Good growth with dark rose-red pigment.
- Nutrient broth: Clear; sediment is formed.
- Chemo-organotrophic: Metabolism is respiratory.
- Acid and gas are not produced from glucose or other carbohydrates in standard medium (method of Subcommittee [19]).
- Catalase is produced (100%).
- Porphyrin respiratory enzymes are produced (100%) (method of Deibel and Evans [5]).
- Acetylmethylcarbinol is not produced.
- β-Galactosidase (*o*-nitrophenyl-β-D-galactopyranoside test) is positive.
- Methyl red is negative.
- Indole and hydrogen sulfide are not produced.
- Nitrate is not reduced.
- Gelatin is hydrolyzed (100%) (method of Clarke [3]).
- Starch is hydrolyzed (90%).
- Esculin may be hydrolyzed (100%).
- Oxidase is produced (100%) (method of Gaby and Hadley [7]).
- Simmons’ citrate: No growth.
- Arginine dihydrolase, ornithine, and lysine decarboxylase and phenylalanine deaminase are not produced.
- Tween 80 may be split (50%).
- Deoxyribonuclease may be produced.

**Table 2. Biochemical characteristics of strains of *Micrococcus agilis***

<table>
<thead>
<tr>
<th>Strain CCM no.</th>
<th>mol% G+C*</th>
<th>Starch hydrolysis</th>
<th>Deoxyribonuclease</th>
<th>Esculin hydrolysis</th>
<th>Tween 80</th>
</tr>
</thead>
<tbody>
<tr>
<td>2539</td>
<td>67.0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>2131</td>
<td>68.4</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>2390</td>
<td>69.0</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>2687</td>
<td>NT</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>2688</td>
<td>NT</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

* All strains tested positive to catalase, benzidine, oxidase and β-galactosidase (*o*-nitrophenyl-β-D-galactopyranoside) and hydrolyzed gelatin. None of the strains produced acid from glucose, lactose, or mannitol, produced acetoin, reduced nitrate, produced phosphatase, urease, tyrosinase, indole, hydrogen sulfide, arginine dihydrolase, ornithine, and lysine decarboxylase, phenylalanine deaminase, or hemolysis, or grew on Simmons’ citrate agar or on nutrient agar with 5% NaCl.
* Data from Boháček et al. [2] and Kocur and Boháček (unpublished data).
* Reaction: +, Positive; -, negative; +, weak; NT, not tested.
Urease, tyrosinase, and phosphatase are not produced.

$\beta$-Hemolysis is not produced.

Aerobic.

Good growth between 20 to 30°C. No growth at 37°C.

No growth on medium with 5% NaCl.

Susceptibility to antibiotics: Susceptible to penicillin, streptomycin, chloramphenicol, tetracycline, erythromycin, novobiocin, ampicillin, carbenicillin, gentamycin, and lysozyme.

Saprophytic.

Habitat: Water, soil, human skin.

G+C content of the DNA is 67.0 to 69.0 mol% (2).

Strain ATCC 966 (= NCTC 7509 = NCDO 983 = CCM 2390) is designated here as the neotype strain of *M. agilis*. A description of this strain follows.

Spheres 0.9 to 1.1 μm in diameter, occurring in pairs and tetrads. Motile by means of one or two flagella. Nonsporeforming. Gram positive.

The cell wall contains peptidoglycan of the L-Lys-L-Thr-L-Ala type.

Agar colonies: Circular, entire, 2 to 3 mm in diameter, slightly convex, smooth, matted, dark rose-red pigmented.

Agar slant: Good growth with dark rose-red, water-insoluble pigment.

Nutrient broth: No turbidity, only sediment is formed.

Chemo-organotrophic: Metabolism is respiratory.

Strictly aerobic.

G+C content of the DNA (69.0 mol%) and biochemical characteristics of the strain are given in Table 2.

Susceptibility to antibiotics: Susceptible to penicillin, streptomycin, chloramphenicol, tetracycline, erythromycin, ampicillin, carbenicillin, gentamycin, novobiocin, and lysozyme. *M. agilis* can be distinguished from the other

\[ \text{G} \rightarrow \text{M} \rightarrow \text{G} \]

\[ \text{L-Ala} \]

\[ \text{d-Glu-NH}_2 \]

\[ \text{L-Lys} \rightarrow \text{L-Thr} \rightarrow \text{L-Ala} \rightarrow \text{L-Ala} \rightarrow \text{d-Ala} \]

\[ \text{d-ALA} \]

\[ \text{L-Lys} \]

\[ \text{d-Glu-NH}_2 \]

\[ \text{L-Ala} \]

\[ \text{G} \rightarrow \text{M} \rightarrow \text{G} \]

**Fig. 2.** Fragment of the primary structure of the peptidoglycan types of *M. agilis*. Abbreviations: *M*, N-acetylmuramic acid; *G*, N-acetylg glucosamine.

### Table 3. Differentiation of species of the genus *Micrococcus*

<table>
<thead>
<tr>
<th>Species*</th>
<th>Pigment</th>
<th>Growth on 1.5% NaCl agar</th>
<th>Growth at 37°C</th>
<th>Glucose (fermentative)</th>
<th>Glucose (aerobic)</th>
<th>Gelatin hydrolysis</th>
<th>Nitrate reduction</th>
<th>Acetoin</th>
<th>Arginine dianhydrolase</th>
<th>Arginase</th>
<th>Reducing</th>
<th>ONPG testa</th>
<th>Motility</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>M. luteus</em></td>
<td>Yellow</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
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</tr>
<tr>
<td><em>M. lylae</em></td>
<td>None</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
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<td>-</td>
<td>-</td>
</tr>
<tr>
<td><em>M. varians</em></td>
<td>Yellow</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td><em>M. roseus</em></td>
<td>Pink</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>d</td>
<td>+</td>
<td>+</td>
<td>-</td>
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<td>-</td>
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<tr>
<td><em>M. agilis</em></td>
<td>Red</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
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<td>-</td>
<td>-</td>
</tr>
<tr>
<td><em>M. kristinae</em></td>
<td>Pale orange</td>
<td>-</td>
<td>+</td>
<td>d</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td><em>M. nishinomiyaensis</em></td>
<td>Orange</td>
<td>-</td>
<td>-</td>
<td>d</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
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</tr>
<tr>
<td><em>M. sedentarius</em></td>
<td>Cream white</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
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</tr>
<tr>
<td><em>M. halobius</em></td>
<td>None</td>
<td>+</td>
<td>-</td>
<td>-</td>
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</tr>
</tbody>
</table>

* For further details, see references 9 and 10.

*ONPG, o-Nitrophenyl-$\beta$-D-galactopyranoside.

* Reaction: +, Positive; -, negative; d, different.

* Requires at least 5% NaCl in the medium. Its description is based on characteristics of single strain only (16).
species of the genus *Micrococcus* by means of characteristics mentioned in Table 3. It is apparent that *M. agilis* differs from *M. roseus* in eight characters, which supports its recognition as a separate species.

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**REPRINT REQUESTS**

Address reprint requests: Dr. M. Kocur, Czechoslovak Collection of Microorganisms, 662 43 Brno, CSSR.

**LITERATURE CITED**